

## MITOCHONDRIAL DNA REVEALS A DIVERGENT PHYLOGENY IN TROPICAL *CENTRUROIDES* (SCORPIONES: BUTHIDAE) FROM MEXICO

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**ABSTRACT.** - Numerous species of highly toxic *Centruroides* scorpions inhabit the tropics of Mexico. Relationships within two common species *C. infamatus* (C. L. Koch, 1844) and *C. limpidus* (Karsch, 1879), and their subspecies, are not known; their existing taxonomy is based on a few morphological characters. In order to further clarify the status of these taxa, we initiated a molecular survey applying mitochondrial (16S rRNA mtDNA sequences) gene markers. Our pilot molecular data set recovers a clear divergent phylogeny among the populations of these species from Michoacán, Guerrero, and Querétaro. Four deep monophyletic clades (putative species) are revealed, with monophyly of *C. limpidus* is not confirmed; *C. limpidus tecomanus* Hoffmann, 1932 does not appear to belong to *C. limpidus*. Phylogenetic, taxonomic, and biogeographic implications are discussed.

**KEY-WORDS.** - Scorpion, Mexico, Michoacán, *Centruroides infamatus*, *C. limpidus*, mtDNA

**RESUME.** - Des nombreuses espèces très toxiques de scorpions du genre *Centruroides* habitent les régions tropicales du Mexique. Les relations entre deux espèces communes, *C. infamatus* (C. L. Koch, 1844) et *C. limpidus* (Karsch, 1879) et leurs sous-espèces ne sont pas connues ; la taxonomie existante est fondée sur quelques caractères morphologiques. Dans le but de clarifier le statut de ces taxa, nous avons débuté une analyse moléculaire avec utilisation de marqueurs géniques mitochondriaux (séquences 16S rARN mtADN). Nos données moléculaires pilotes démontrent une nette divergence phylogénétique entre les populations de ces espèces, originaires de Michoacán, Guerrero et Querétaro. Quatre clades monophylétiques (espèces putatives), sont révélées, avec la monophylie de *C. limpidus* non confirmée ; *C. limpidus tecomanus* Hoffmann, 1932 ne semble pas appartenir à *C. limpidus*. Des implications taxonomiques et biogéographiques sont discutées.

**MOTS-CLES.** - Scorpion, Mexico, Michoacán, *Centruroides infamatus*, *C. limpidus*, mtADN

### INTRODUCTION

*Centruroides* Marx, 1890, with 41 species and 24 subspecies (FET & LOWE, 2000) is one of the most diverse genera of the scorpion family Buthidae (second only to *Tityus* C. L. Koch), and the only genus of this family found in North America. Scorpions of this genus (including many Central Mexican species) are among the most toxic in the world for humans and other mammals (MAZZOTTI & BRAVO-BECHERELLE, 1963; SIMARD &

WATT, 1990). They are abundant in various natural habitats ranging from tropical forest to temperate deserts. *Centruroides* is especially diverse in Mexico and the Caribbean, but is also found in Central and South America (SISSOM & LOURENÇO, 1987; FET & LOWE, 2000). Taxonomy of many *Centruroides* species is confusing, and was traditionally based mainly or exclusively on the features of morphosculpture and coloration characters. No modern taxonomic or phylogenetic analysis exists for this genus, and the only existing generic key (STAHNKE & CALOS, 1977) is outdated. The only faunal treatment of the Mexican fauna was that by HOFFMANN (1932). The keys and lists for 32 species and subspecies of *Centruroides* found in Mexico were produced by DÍAZ NÁJERA (1964, 1966, 1975), BEUTELSPACHER (2000); FET and LOWE (2000). Although a comparative study (DEHESA-DÁVILA *et al.*, 1996) on the primary structures of Na<sup>+</sup> channel-specific toxins in *C. infamatus infamatus* (C. L. Koch, 1844) and in *C. limpidus limpidus* (Karsch, 1879) has recently been done, the phylogenetic relationships among these species remain uncertain.

LOURENÇO and SISSOM (2000) reviewed scorpion diversity in Mexico, emphasizing the need for more investigation and novel approaches (including DNA techniques) to understand complex taxonomy, origin, and distribution of Mexican *Centruroides*. They say (p. 117): "...it is likely that some of these species will prove to be synonyms, and equally likely that others will represent complexes of sibling species rather than single species".

This paper is a part of an ongoing study of scorpion fauna, ecology, biology, and biogeography in the state of Michoacán (PONCE & BEUTELSPACHER, 2001). We report here the first pilot data based on the analysis of mitochondrial DNA (mtDNA) sequences of the 16S rRNA gene from two common species of *Centruroides* endemic to Central Mexico. These molecular markers have recently become a powerful tool for evaluating the taxonomic status of animal populations, subspecies, and species. The first information on applicability of mtDNA analysis to the species-level taxonomy of *Centruroides* was reported by FET and POINDEXTER (1992). Recently, comparisons of 16S rRNA mtDNA sequences allowed to clarify phylogeny at the species level among the populations of *Centruroides exilicauda* (Wood, 1863) from Baja California and Sonora (Mexico) and Arizona (USA) (FET *et al.*, 1999; GANTENBEIN *et al.*, 2001) as well as other scorpion genera and families (FET *et al.*, 2001; GANTENBEIN *et al.*, 1999, 2000). The number of base pairs that are sequenced from this gene fragment was high enough for a good phylogenetic signal (HUELSENBECK & HILLIS, 1996).

## MATERIAL AND METHODS

### SPECIMENS ANALYSED

Total of 14 specimens of two species of *Centruroides* were analysed; each was represented by its two existing subspecies. These specimens were collected and identified by J.P.S. from the following 12 localities in Mexico (see Fig. 1; abbreviations as given below): *Centruroides infamatus infamatus* (C. L. Koch, 1844): Zumpimito, 1560 m a.s.l., Municipio de Uruapan, Michoacán, Mexico (CiiZU1). – *C. infamatus ornatus* Pocock, 1902 (all Michoacán, Mexico): El Cobano, 1540 m, Municipio de Gabriel Zamora (CioEC1); La Caratagua, 1680 m, Municipio de Coeneo (CioLC1); near Morelia city, 1940 m, Municipio de Morelia (CioMO1); Tiripetio, 2020 m, Municipio de Morelia (CioTI1). – *C. limpidus limpidus* (Karsch, 1879): Churumuco, 300 m, Michoacán, Mexico (CiiCH1, CiiCH2); Arúa, 480 m, Municipio de Huetamo, Michoacán, Mexico (CiiAR1); Tzirandaro, 500 m, Guerrero, Mexico (CiiTO1); Tres Puentes, Municipio de

Tzitzio, Michoacán, Mexico, 1775 m (CITZ1); Huitzucó, 960 m, near Iguala, Guerrero, Mexico (CIHU1); Cañon de los Cajones, 1820 m, Municipio de Querétaro, Querétaro, Mexico (CIQU1). – *C. limpidus tecomanus* Hoffmann, 1932: El Faro de Buceras, Municipio de Coahuayana, Michoacán, Mexico (ClFB1, ClFB2). These localities represented a broad geographic spectrum of habitats and climates characteristic for the study area, and varied in altitude from 300 to 1940 m a.s.l.

Three congeneric species were used for comparison, *C. exilicauda* (Wood, 1863) from Tempe, Arizona, USA (coll. J. Bigelow) (CexTE), *C. vittatus* (Say, 1821) from Arkansas, USA (coll. T. Yamashita) (CviAR), and *C. bani* Armas & Marcano Fondeur, 1987 from Dominican Republic (coll. D. Huber) (CbaDO). Animals were shipped on dry ice to Marshall University (West Virginia, USA) and preserved in 96 % ethanol for DNA analysis. Voucher specimens are deposited in V.F.'s collection at Marshall University.

## DNA EXTRACTION AND SEQUENCING TECHNIQUES

Comparative analysis of the mitochondrial (mt) 16S ribosomal RNA is widely used in modern molecular evolutionary studies of various arthropods (SIMON *et al.*, 1994). For detailed techniques and protocols see GANTENBEIN *et al.* (1999, 2000). Total DNA was extracted from preserved (in 95 % ethanol) muscle tissue using a standard extraction method (Qiagen™). An approximately 400 bp fragment of the 16S rRNA gene was amplified by the polymerase chain reaction (PCR) using the universal forward primer LR-J-12887 (SIMON *et al.*, 1994) and a scorpion-specific reverse primer (GANTENBEIN *et al.*, 1999).

## PHYLOGENETIC ANALYSES

17 mtDNA sequences representing different haplotypes (including ambiguities) were aligned using Clustal X (HIGGINS *et al.*, 1991) and by eye. All ambiguities and gaps were omitted for further phylogenetic analyses as described in SWOFFORD *et al.* (1996), resulting in 366 characters, and 15 unique haplotypes, which were deposited in the GenBank database. For the reconstruction of phylogenetic relationships we applied Maximum Likelihood (FELSENSTEIN, 1981) analysis and Maximum Parsimony to our DNA data. In order to select the most appropriate DNA model of nucleotide substitution we calculated hierarchic likelihood ratio test statistics using the program Modeltest 3.06 Posada which is implemented in PAUP\* 4.0b8 (SWOFFORD, 1998) and tests 56 different substitution models based on a NJ tree using Jukes-Cantor (JUKES & CANTOR, 1969) distances. Details about likelihood ratio tests are given in HUELSENBECK and RANNALA (1997) and in HUELSENBECK and CRANDALL (1997). The HKY85 +  $\Gamma$  model (HASEGAWA *et al.*, 1985) was selected. The rate heterogeneity among sites was assumed to follow a gamma distribution (shape parameter  $\alpha$  was ML-estimated) with four categories, each represented by its mean (YANG, 1996). In a further step, the molecular clock hypothesis (i.e., equal rates across all sequences) was tested using the  $\chi^2$  approximated likelihood ratio test statistics with OTUs-2 degrees of freedom ( $df = 17$  minus 2 = 15) which was not rejected with a  $P$ -value of 0.09. Therefore, we explored the tree space by 100 heuristic tree searches and by randomising the order of the sequence input using the clock enforcement option in PAUP\*. Transitions (ti) were down-weighted relative to transversions (tv) according to the ML-estimated ti/tv ratio, which was about 7:1 in favour of ti (see Results). Tree stability of best trees was evaluated by calculating the consistency index excluding uninformative sites (CIu), and the retention index (RI) (KITCHING *et al.*, 1998). We also applied the genetic distance approach, i.e. the comparison of pairwise genetic distances between haplotypes. We used both the

uncorrected sequence divergence and the corrected divergence using the HKY+ $\Gamma$  model with the parameters estimated from ML.

In all phylogenetic trees *Centruroides vittatus* (Say, 1821) and *C. bani* Armas & Marcano Fondeur, 1987 were preset as outgroups. *C. exilicauda* (Wood, 1863) was tested as an outgroup to *C. limpidus* and *C. infamatus*, and thereafter treated as an ingroup. All tree searches were performed by the branch-and-bound algorithm. Reliability of inferred topologies of trees were assessed using the bootstrap procedure (FELSENSTEIN, 1985), and resampled 1,000 pseudo-replicates except for ML where we resampled 100 pseudo-replicates because of computational time.

## DNA SEQUENCE AVAILABILITY

All 14 new sequences were deposited to the GenBank Nucleotide Sequence Database with the following accession numbers: *C. infamatus infamatus*: CiiZU1 = AF439753; *C. infamatus ornatus*: CioEC1 = AF439754; CioLC1 = AF439755; CioMO1 = AF439756; CiiTI1 = AF439757; *C. limpidus limpidus*: CiiCH1 = AF439758; CiiCH2 = AF439759; CiiAR1 = AF439760; CiiTO1 = AF439761; CiiTZ1 = AF439762; CiiHU1 = AF439763; CiiQU1 = AF439764; *C. limpidus tecomanus*: CltFB1 = AF439765; CltFB2 = AF439766. Three outgroup sequences were published previously (GANTENBEIN et al., 2001): *C. exilicauda*: CexTE = AJ288639; *C. vittatus*: CviAR = AJ288643; *C. bani*: CbaDO = AJ288644

## RESULTS

The calculated pairwise genetic distances between the 17 sequences are presented in Table I.

The two tree topologies inferred from Maximum Likelihood and Maximum Parsimony (Fig. 2) are almost congruent except for the phylogenetic position of the *Centruroides exilicauda* sequence. The analyses revealed four well supported main clusters, with the following results:

(a) Monophyly of *C. infamatus* was confirmed, with a sole *C. infamatus infamatus* specimen from Zumpimito (CiiZU1) forming a sister clade to all four sampled populations of *C. infamatus ornatus* (CioCO1, CioLC1, CioTZ1, CioTI), which all grouped closely together.

(b) The subspecies *C. limpidus tecomanus* was represented in our samples by two specimens from El Faro de Bucerías (CltFB1, CltFB2) at the coastal zone of southwestern Michoacán. Two close DNA haplotypes from this population did not form a monophyletic group with specimens of *C. limpidus limpidus*, but instead formed an unexpected but highly supported sister clade to *C. infamatus*. Therefore, monophyly of *C. limpidus* was not confirmed.

(c) Within the seven studied specimens of *C. limpidus limpidus*, two deeply divergent, well-supported clades were observed. One clade was represented by four populations at the Balsas Depression area in Michoacán (CiiCH1, CiiCH2, CiiAR1, CiiTZ1, CiiMO1) which grouped closely together. The second clade recovered within *C. limpidus limpidus* included two specimens from two remote mountainous areas further northwest (Querétaro, CiiQU1) and southwest (Guerrero, CiiHU1).

## DISCUSSION

*Centruroides* is one of a few extant scorpion genera for which Cenozoic fossils exist, including the Miocene/Oligocene amber of Chiapas, Mexico, with minimum age 22.5 - 26 million years (Ma) (SANTIAGO-BLAY & POINAR, 1993). Thus, the age of the extant lineages can be very old, and their evolution could be considered against the geological events. GANTENBEIN *et al.* (2001) demonstrated recently for *Centruroides* from Sonora and Baja California, that speciation events in the deserts of northwestern Mexico could be of at least the Miocene age (5-3 Ma). For 16S rRNA gene in *Centruroides*, the evolutionary rate (sequence divergence per time unit; LI, 1997) was estimated to vary from 1.4-2.3% uncorrected sequence divergence ( $p$ ) and from 3-5% HKY +  $\Gamma$  distance per Ma (GANTENBEIN *et al.*, 2001). These estimates of molecular evolutionary rates in mitochondria lie in the range of the clock calibration in primates by BROWN *et al.* (1979) based on mtDNA RFLP data. This study predicts an evolutionary rate of mitochondrial DNA of about 2% uncorrected divergence per Ma and was cited many times for organisms where no fossil data or dated vicariant events are available. More important for invertebrates, however, is the estimated divergence rate for the mitochondrial 16S gene in Hawaiian *Drosophila* species that was again estimated to about 2% uncorrected divergence per Ma (DESALLE *et al.*, 1987). Another clock calibration used pairwise genetic divergences of sister taxa of snapping shrimps of the genus *Alpheus* across the Isthmus of Panama and revealed a rate of 1.4% corrected sequence divergence for the mitochondrial cytochrome oxidase I gene (KNOWLTON & WEIGHT, 1998). These rates imply that the divergence rate in the scorpion mitochondrial 16S rRNA gene is comparable to other rates estimated in other invertebrates and even in vertebrates although scorpions have a much longer (mammalian-like) generation time (~2 yrs, for buthids; POLIS and SISSOM, 1990) compared to *Drosophila* sp. (~14 days), and have one of the lowest known metabolism rates in the animal kingdom (HADLEY, 1990). Consequently, we expect a lower rate in scorpion genes than in genes of other animal groups assuming that variation in evolutionary rates across organism groups is mainly due to differences in generation time or metabolism rate (MARTIN & PALUMBI, 1993).

Our pilot molecular data set analysis, first of all, confirmed monophyly of the studied populations of *Centruroides infamatus*. There were clearly observed two clade within this species. One was represented by the sole *C. infamatus infamatus* specimen taken from a humid pine-oak forest of Zumpimito, on the southern slope of the Trans-Mexican Volcanic Belt mountain range (these mountains are also known as "Corredor Tarasco"). Another clade included four specimens identified as *C. infamatus ornatus* collected from the same landscape as Zumpimito (El Cobano, 1540 m) and three colder and drier highland localities (La Caratacua, 1680 m; Morelia city, 1940 m; and Tiripetio, 2020 m) of the northern slope. The taxonomic status of these two subspecies is unresolved. They were originally described as separate species, and *C. ornatus* (type locality Jalisco, Mexico) was later downgraded to a subspecies of *C. infamatus* by HOFFMANN (1932). Most recently, BEUTELSPACHER (2000) synonymized this taxon with *C. infamatus* (treating it as a "forma *ornatus*") is not in accordance with the International Code of Zoological Nomenclature, which does not allow categories below subspecies rank). However, PONCE and BEUTELSPACHER (2001) suggested that these two subspecies do in fact have certain morphometric differences; new preliminary data support this conclusion (Table II). Moreover, at least in Michoacán these two forms were never found sympatrically (observations of J.P.S.; the data from literature showing sympatry of *C. i. infamatus* and *C. i. ornatus* are based on misidentifications). Our pilot DNA data confirm two divergent clades; divergence rate estimates their common ancestor's age as 2-4 Ma for HKY+ $\Gamma$  divergence rate ( $11.7 \pm 0.9$  %) and 3-5 Ma for uncorrected  $p$  ( $7.2 \pm 0.4$  %). Further study is necessary with sampling all over the range of both taxa, to confirm existence of two independent lineages. If such are confirmed, *C. i. ornatus* could be elevated to a species rank. In our opinion the nomenclatural category

of subspecies is not consistent with the viewpoint of the phylogenetic species concept (PSC), which defines species based on monophyly.

For *Centruroides limpidus*, the situation was more complicated, since monophyly of this species was not confirmed by the DNA data. The coastal population from El Faro de Bucierias, identified as the subspecies *C. l. tecomanus*, formed an unexpected but highly supported sister clade to *C. infamatus* rather than to *C. l. limpidus*! Moreover, at least in one of our analyses *C. exilicauda* from Arizona formed an ingroup, cutting between *C. l. limpidus* clade and *C. limpidus tecomanus* (Fig. 2). The evolutionary rate estimate of the age of common ancestor of *C. infamatus* and "*C. limpidus tecomanus*" is ca. 3-5 Ma for HKY+ $\Gamma$  divergence rate ( $15.3 \pm 1.9\%$ ), and 4-7 Ma for uncorrected  $p$  ( $10.1 \pm 0.8\%$ ).

We thus suspect that "*C. l. tecomanus*" is, in fact, a separate species, which may not be closely related to *C. limpidus* at all. This enigmatic taxon was not studied in detail since its description by HOFFMANN (1932) from several localities in the Mexican state of Colima (Colima, Tecoman, and Manzanillo). It is differentiated from the nominotypic subspecies essentially only by presence of a developed subaculear tooth (STAHNKE & CALOS, 1977) – a character which is known to vary even within developmental stages in *Centruroides* (LOURENÇO, 1982); some juvenile specimens identified formerly as *C. l. tecomanus* appear in fact to be juveniles of *C. l. limpidus* (PONCE & BEUTELSPACHER, 2001). Additional morphological data (PONCE, in preparation) show that this form also is distinguished from *C. limpidus limpidus* by the shape of female pectinal teeth, the color pattern on the carapace, and the carination of mesosomal tergites. The recent comparative study by DEHESA-DÁVILA *et al.* (1996) on the primary amino acid structure of the Na<sup>+</sup>-channel-specific  $\beta$ -toxins supports our view that *C. l. tecomanus* is highly divergent from *C. limpidus*. Comparing the amino acid sequences, in their study, *i.e.* the *Cii-1* toxin (= *C. i. infamatus*) and two *Cll* toxins (= *C. l. limpidus*) with the *Clt-1* toxin (= *C. l. tecomanus*) revealed that 3-4% amino acid replacements were found between the *Cii* and the *Cll* sequences, whereas 4-7% substitutions were found between *Clt-1* and all other toxins. Moreover, a phylogenetic tree reconstructed from their alignment by Maximum Parsimony (tree not shown) revealed that *Clt-1* clearly was found separated from the two *Cll1-2* and *Cii-1* sequences. Although these sequences might represent actually paralogous genes of a closely related toxin gene family, our phylogenetic approach indicates that toxin genes harbor considerably phylogenetic information despite the high selective constraint. In this context, especially synonymous mutations (expected to be neutral) at the DNA level would be more informative since amino acid replacements might be of adaptive nature. The geographic range of *C. l. tecomanus* is confined to a rather narrow ecological area along the Pacific coast (Región de la Costa) within the states of Nayarit, Colima, Michoacán, and Jalisco. *C. l. tecomanus* does not cross the mountains of Sierra Madre del Sur into the Región de Sierras del Centro in Michoacán (PONCE & BEUTELSPACHER, 2001), and thus is practically allopatric with *C. l. limpidus*.

The nominotypic subspecies *C. limpidus limpidus* has a rather wide range in the south-central Mexico (central Guerrero, Morelos, Michoacán, Distrito Federal, Mexico, Querétaro, and parts of Oaxaca and Puebla). We sampled its populations from Michoacán, Guerrero, and Querétaro. Within this material, the DNA phylogeny reveals two clear lineages of deep divergence. One lineage (well supported at 79 to 84% bootstrap) includes specimens from Churumuco, Arua, Tzirandaro, and Tzitzio, all found within the boundaries of the Balsas Depression in Michoacán, along the entire altitudinal profile (300 to 1775 m a. s. l.). Another well-supported lineage (96 to 97% bootstrap) includes two specimens sampled from the mountains further to the west (two distant localities in Querétaro and Guerrero). Time of divergence between two lineages can be estimated at ca. 5-8 Ma BP for both HKY+ $\Gamma$  divergence ( $23.7 \pm 3.5\%$ ) and uncorrected  $p$  ( $10.9 \pm 1.0\%$ ) (Fig. 2). This estimate is consistent with the geological age of Balsas Depression, which was formed from 4 to 6 Ma BP. The mountains of Sierra Madre del

Sur in the south, and the Mexican Transverse Volcanic Belt in the north and west have been rising since the Eocene-Oligocene time. The tectonic events could have isolated the populations in the area of modern Balsas Depression, which possibly could speciate *in situ* and then disperse toward the adjacent mountain slopes of the transitional zone (e.g. Tzitzio, 1775 m). The divergence time between Querétaro and Guerrero samples was estimated as 2-4 Ma BP for HKY+ $\Gamma$  rate (11 %) and 3-5 Ma for uncorrected  $p$  (7 %).

Our results, therefore, suggest that more than one ancient monophyletic lineage (possibly, more than one species) exist within currently accepted *Centruroides limpidus limpidus*. Moreover, our preliminary morphological data indicate that there is some difference between two clades identified by this DNA analysis. The number of the setae on the anterior margin of the carapace differs significantly between the Balsas Depression populations (8-11, usually 8 with 4 on each side,  $n=29$ ) from that in Querétaro and Huiztoco, Guerrero (6-7 setae, usually 6 with 3 on each side,  $n=13$ ). The type locality of *Centruroides limpidus* is Puebla, which lies in the same geographic area as Guerrero. Thus, we might assume that the Querétaro/Guerrero lineage corresponds to "true" *C. limpidus*, and that the Balsas Depression populations could belong to another, "cryptic", or "sibling" species. Our DNA analyses revealed similar situations in *C. exilicauda* (GANTENBEIN *et al.*, 2001) and in the Alpine scorpion species *Euscorpium germanus* (C.L. Koch) (Euscorpiidae) (GANTENBEIN *et al.*, 2000). Local speciation and endemism are common in the studied area, which forms a biogeographic boundary for many faunal groups between Nearctic and Neotropics (FA & MORALES, 1993). Further, detailed investigations should be done to test our preliminary conclusions: one has to study many more populations from the entire range of *C. limpidus*. Several data sets (mitochondrial and nuclear genes, allozymes, morphology, toxin structure/activity, etc.) could be analysed to establish the true taxonomic and genetic structure of the populations and species of *Centruroides*.

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As LOURENÇO & SISSOM (2000, p. 124) stated, Mexico has a significant tradition in the study of scorpions, related to the important public health problems due to scorpion stings, and "training of Mexican specialists and/or collaborative research between scientists in Mexico and those of other countries would be highly desirable". We hope that our international collaboration will be one of the steps toward this goal.

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Table I. Maximum Likelihood distance matrix (upper right) and uncorrected  $p$  distance matrix (lower left) of *Centruroides* 16S mtDNA sequences. Parameters for ML-distances were set to the HKY85 +  $\Gamma$  model; transition/transversion ratio = 7.44 ( $\kappa$  = 18.41); assumed nucleotide frequencies (set by user): A=0.35, C=0.15, G=0.12, T=0.38; shape parameter  $\alpha$  = 0.15. 58 characters are excluded, 366 characters remaining.

|                  | 1    | 2    | 3    | 4    | 5    | 6    | 7    | 8    | 9    | 10   | 11   | 12   | 13   | 14   | 15   | 16   | 17   |
|------------------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| 1 <i>CioLC1</i>  | -    | 0.00 | 0.00 | 0.01 | 0.11 | 0.14 | 0.16 | 0.22 | 0.23 | 0.21 | 0.22 | 0.22 | 0.50 | 0.41 | 0.20 | 0.28 | 0.37 |
| 2 <i>CioEC1</i>  | 0.00 | -    | 0.00 | 0.01 | 0.11 | 0.14 | 0.16 | 0.22 | 0.23 | 0.21 | 0.22 | 0.22 | 0.50 | 0.41 | 0.20 | 0.28 | 0.37 |
| 3 <i>CioMO1</i>  | 0.00 | 0.00 | -    | 0.01 | 0.12 | 0.15 | 0.16 | 0.22 | 0.22 | 0.22 | 0.23 | 0.23 | 0.51 | 0.42 | 0.21 | 0.29 | 0.36 |
| 4 <i>CioT11</i>  | 0.01 | 0.01 | 0.01 | -    | 0.13 | 0.16 | 0.19 | 0.25 | 0.26 | 0.24 | 0.25 | 0.25 | 0.54 | 0.42 | 0.22 | 0.31 | 0.38 |
| 5 <i>ChiZU1</i>  | 0.07 | 0.07 | 0.07 | 0.08 | -    | 0.12 | 0.14 | 0.21 | 0.22 | 0.20 | 0.24 | 0.24 | 0.53 | 0.46 | 0.23 | 0.29 | 0.40 |
| 6 <i>ChiFB1</i>  | 0.08 | 0.08 | 0.08 | 0.09 | 0.07 | -    | 0.01 | 0.22 | 0.23 | 0.21 | 0.20 | 0.20 | 0.41 | 0.40 | 0.20 | 0.28 | 0.36 |
| 7 <i>ChiFB2</i>  | 0.09 | 0.09 | 0.09 | 0.10 | 0.08 | 0.01 | -    | 0.22 | 0.23 | 0.24 | 0.23 | 0.23 | 0.44 | 0.44 | 0.23 | 0.31 | 0.35 |
| 8 <i>ChiCH1</i>  | 0.10 | 0.10 | 0.10 | 0.11 | 0.10 | 0.10 | 0.09 | -    | 0.00 | 0.03 | 0.08 | 0.08 | 0.54 | 0.39 | 0.19 | 0.20 | 0.23 |
| 9 <i>ChiCH2</i>  | 0.11 | 0.11 | 0.10 | 0.11 | 0.10 | 0.10 | 0.10 | 0.00 | -    | 0.03 | 0.09 | 0.09 | 0.56 | 0.40 | 0.18 | 0.21 | 0.22 |
| 10 <i>ChiTO1</i> | 0.10 | 0.10 | 0.10 | 0.11 | 0.09 | 0.09 | 0.10 | 0.02 | 0.03 | -    | 0.06 | 0.06 | 0.46 | 0.40 | 0.20 | 0.19 | 0.26 |
| 11 <i>ChiTZ1</i> | 0.10 | 0.10 | 0.10 | 0.11 | 0.10 | 0.09 | 0.10 | 0.05 | 0.06 | 0.04 | -    | 0.00 | 0.44 | 0.40 | 0.23 | 0.24 | 0.29 |
| 12 <i>ChiAR1</i> | 0.10 | 0.10 | 0.10 | 0.11 | 0.10 | 0.10 | 0.09 | 0.05 | 0.06 | 0.04 | 0.00 | -    | 0.44 | 0.40 | 0.23 | 0.24 | 0.29 |
| 13 <i>CbaDO</i>  | 0.14 | 0.14 | 0.14 | 0.15 | 0.14 | 0.13 | 0.13 | 0.14 | 0.14 | 0.12 | 0.12 | 0.12 | -    | 0.46 | 0.40 | 0.50 | 0.60 |
| 14 <i>CviAR</i>  | 0.15 | 0.15 | 0.15 | 0.15 | 0.15 | 0.14 | 0.15 | 0.14 | 0.14 | 0.14 | 0.14 | 0.14 | 0.14 | -    | 0.36 | 0.49 | 0.38 |
| 15 <i>CexTE</i>  | 0.10 | 0.10 | 0.11 | 0.11 | 0.11 | 0.10 | 0.11 | 0.09 | 0.09 | 0.10 | 0.11 | 0.11 | 0.13 | 0.14 | -    | 0.29 | 0.27 |
| 16 <i>ChiQU1</i> | 0.12 | 0.12 | 0.13 | 0.13 | 0.12 | 0.12 | 0.13 | 0.10 | 0.10 | 0.09 | 0.11 | 0.11 | 0.15 | 0.16 | 0.12 | -    | 0.11 |
| 17 <i>ChiHU1</i> | 0.15 | 0.15 | 0.15 | 0.16 | 0.15 | 0.14 | 0.14 | 0.11 | 0.11 | 0.12 | 0.12 | 0.12 | 0.18 | 0.14 | 0.12 | 0.07 | -    |

Table II. Morphometric comparison of adult *C. infamatus infamatus* (21 females, 8 males) and *C. i. ornatus* (total 10 females, 10 males).

| Character                          | <i>C. i. infamatus</i> | <i>C. i. ornatus</i>  |
|------------------------------------|------------------------|-----------------------|
| Pectinal tooth number, female      | 21–25 (ave. 22.0)      | 17–22 (ave. 18.5)     |
| Pectinal tooth number, male        | 22–26 (ave. 24.0)      | 20–22 (ave. 21.0)     |
| Movable finger length, female (mm) | 5.60–6.75 (ave. 6.09)  | 4.70–5.80 (ave. 5.13) |
| Movable finger length, male (mm)   | 5.15–6.35 (ave. 5.71)  | 4.25–5.70 (ave. 5.22) |
| Chela width, female (mm)           | 1.90–2.50 (ave. 2.13)  | 1.55–1.95 (ave. 1.72) |
| Chela width, male (mm)             | 1.85–2.30 (ave. 2.01)  | 1.80–1.90 (ave. 1.83) |

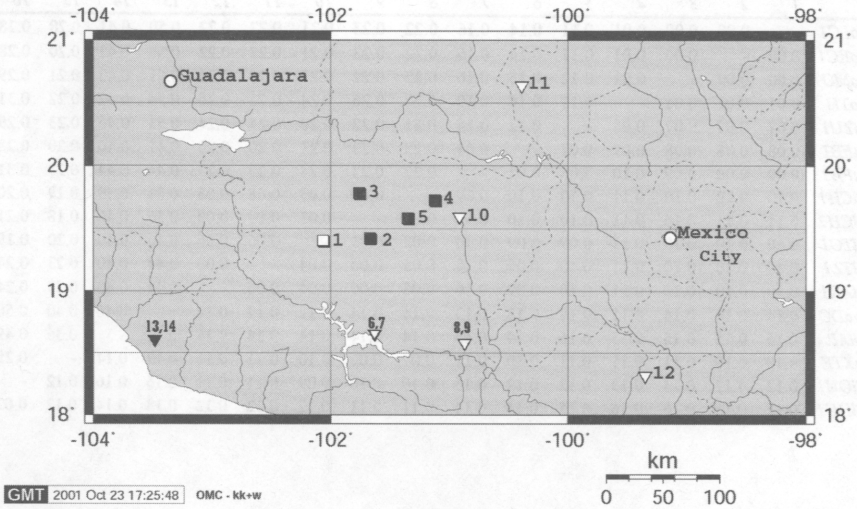


Fig. 1. Map of the samples of *Centruroides* in Central Mexico (see detailed localities data in text). *C. infamatus infamatus*: 1 - CiiZU1, Zumpimito; *C. infamatus ornatus*: 2 - CioCO1, El Cobano; 3 - CioLC1, La Caratacua; 4 - CiiMO1, Morelia; 5 - CioTI1, Tiripetio; *C. limpidus limpidus*: 6 - CII CH1 and 7 - CII CH2, Churumuco; 8 - CII TO1, Tzirandaro; 9 - CII AR1, Arua; 10 - CII TZ1, Tzitzio; 11 - CII QU1, Querétaro; 12 - CII HU1, Huitzuco; *C. limpidus tecomanus*: 13 - CII FB1 and 14 - CII FB2, El Faro de Bucerias.

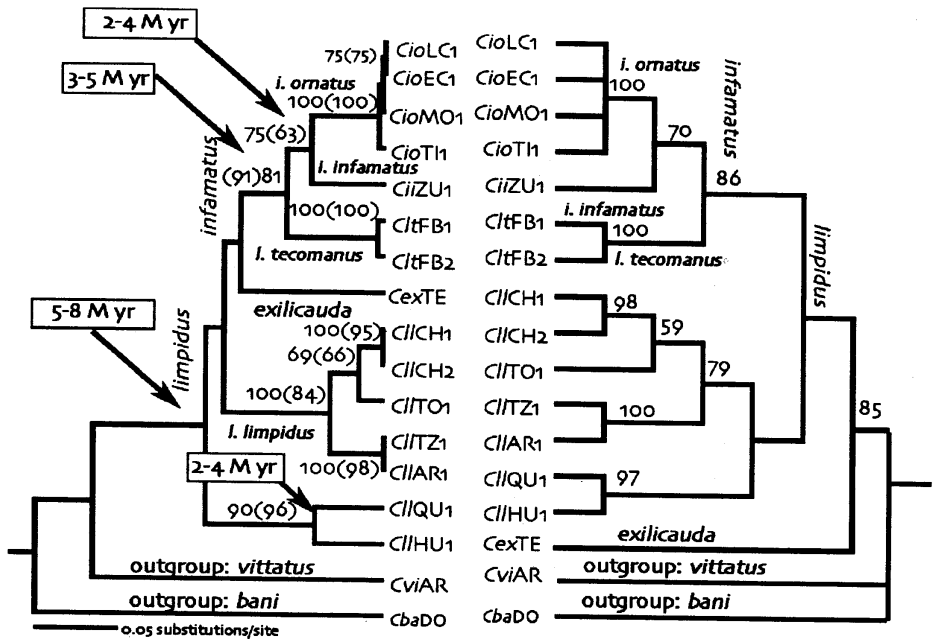


Fig. 2. 16S mtDNA gene genealogy (366 bp) of highly toxic *Centruroides* from central Mexico. Included are the subspecies *C. infamatus infamatus*, *C. i. ornatus*, *C. limpidus limpidus*, and *C. l. tecomanus*. Tree topology was revealed by Maximum Likelihood (ML) analysis (left) using the HKY85 +  $\Gamma$  model and by weighted Maximum Parsimony (MP) analysis (right). The parameters for ML were estimated to  $\pi_A = 0.34$ ,  $\pi_C = 0.15$ ,  $\pi_G = 0.12$ , and  $\pi_T = 0.37$ ,  $\alpha = 0.15$ , transition (ti)/transversion (tv) ratio = 7.45 ( $\kappa = 18.41$ ),  $-\ln L = 1411.76$ . For MP, ti's were down-weighted relative to tv's by factor seven. The single MPtree needed 209 steps, and the Consistency Index excluding uninformative characters (CIu) and the Retention Index (RI) were 0.56 and 0.70, respectively. The tree topology of the cladogram estimated by ML differs from the topology revealed by MP only in the phylogenetic position of the sequence of *C. exilicauda*. Bootstrap values correspond to bootstrapping (in parentheses from Neighbour-Joining analysis). Divergence time estimates are given for ML tree under HKY85 +  $\Gamma$  model (see 'Discussion' for details). Abbreviations for haplotypes are given in Materials & Methods.

Addendum: 16S mtDNA sequence alignment for subspecies *C. infamatus infamatus*, *C. i. ornatus*, *C. limpidus limpidus*, and *C. l. tecomanus*.

CioLC1 NNNNNAGNCCACTTTACAGGTCGGAACAGACCTCCTTTNACTCCTCTTG  
 CioEC1 NNNNNNGCCTTTTACAGGTCNGAACAGACCTCCTTTNACTCCTCTTG  
 CioMO1 NNNNNNGGTTTTTTACAGGTCNGAACAGACCTCCTTTNACTCCTCTTG  
 CioTI1 NTNTTGGANCCNNTNAGGACNNGCCAGACCTCCTTTNACTCCTCTTG  
 CiiZU1 NTNTTNNCGCCTNTTTTTGGGTCCGAACAGACCCCTTTTACTCCTCTTG  
 CltFB1 NNNCTTNGCCACTTTATGGGTTCNGAACAGACCCCTTTTACTCCTCTTG  
 CltFB2 NNNNNGGTTTTCTTTATGNGTCNGAACAGNCCNCTTTTACTCCTCTTG  
 Cl1CH1 NNNTGGTTTTATTTATGNGTCNGAACAGNACCCCTTTTATTCCTCTTG  
 Cl1CH2 NNNNNGGTTTTATTTATGNGTCNGAACAGNCCCTTTTATTCCTCTTG  
 Cl1TO1 NNNNNNGNCCANTCATGGGTCCNGAACAGANCCCTTTTATTCCTCTTG  
 Cl1TZ1 NNNNNNGAGCCCTTNTGGGTTCNGNCCAGANCCCTTTTATTCCTCTTG  
 Cl1AR1 TTTTNGAAGCCCTTTATGGGTTCNGNCCAGANCCCTTTTATTCCTCTTG  
 Cl1QU1 NNNNNNGCCCTTTTATGNGTCNGAACAGACCCCTTTTATTCCTCTTG  
 Cl1HU1 NNNNNNGGTCCTTTTATCGNCTCGACAGNACCTTCTTTTATCCCTCTTG  
 CbaDO NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNCCCTATTAATACTCTTG  
 CviAR NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNCTTCTATTACTCTTG  
 CexTE NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNTCCCTCCTCTTG

CioLC1 CGGAGTGAAGGAAATCTAATCCAACATCGAGGTCGCAAACGTATTTGTCA  
 CioEC1 CGGAGTGAAGGAAATCTAATCCAACATCGAGGTCGCAAACGTATTTGTCA  
 CioMO1 CGGAGTGAAGGAAATCTAATCCAACATCGAGGTCGCAAACGTATTTGTCA  
 CioTI1 CGGAGTGAAGGAAATCTAATCCAACATCGAGGTCACAAACGTATTTGTCA  
 CiiZU1 CGGAATAGAGGAAATCTAATCCAACATCGAGGTCGCAAACATATTCGTCA  
 CltFB1 CGGAATAAAGGAAATCTAATCCAACATCGAGGTCGCAAACATATTTGTCA  
 CltFB2 CGGAATAAAGGAAATCTAATCCAACATCGAGGTCGCAAACATATTTGTCA  
 Cl1CH1 CGGAATAGTGGAAATCTAATCCAACATCGAGGTCGCAAACATATTTGTCA  
 Cl1CH2 CGGAATAGTGGAAATCTAATCCAACATCGAGGTCGCAAACATATTTGTCA  
 Cl1TO1 CGGAATAGTGGAAATCTAATCCAACATCGAGGTCGCAAACATATTTGTCA  
 Cl1TZ1 CGGAATAATGGAAATCTAATCCAACATCGAGGTCGCAAACATATTCGTCA  
 Cl1AR1 CGGAATAATGGAAATCTAATCCAACATCGAGGTCGCAAACATATTCGTCA  
 Cl1QU1 CGGAATAGTGGAAAGCTTAATCCAACATCGAGGTCGCAAACATATTTGTCA  
 Cl1HU1 CGGGATAGAGGAAAGCTTAATCCAACATCGAGGTCGCAAACATGTTTGTCA  
 CbaDO CATAATAAAGGAAACTAATCCAACATCGAGGTCGCAAACACATATTTGTCA  
 CviAR CGTAATACAGGGAATTTAATCCAACATCGAGGTCACAAACACGTTTATCA  
 CexTE CGGAGTGGAGGAAACTTAATCCAACATCGAGGTCGCAAACATATTTGTCA

CioLC1 ATTTGAACCTTAAAAATACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CioEC1 ATTTGAACCTTAAAAATACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CioMO1 ATTTGAACCTTAAAAATACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CioTI1 ATTTGAACCTTAAAAATACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CiiZU1 ATTTGAGCTTTAAGAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CltFB1 ATTTGAGCTTTAAAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CltFB2 ATTTGAGCTTTAAAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT  
 Cl1CH1 ATTTGAACCTTTCAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT  
 Cl1CH2 ATTTGGACTTTCAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT  
 Cl1TO1 ATTTGAGCTTTCAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT  
 Cl1TZ1 ATTTGAACCTTTCAAATACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 Cl1AR1 ATTTGAACCTTTCAAATACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 Cl1QU1 ATTTGAGCTTTCAAATACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 Cl1HU1 ATTTGGACTTTCAAACACATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CbaDO ATTTGAACCTTTCAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT  
 CviAR ATTTGAACCTTTCAAACATATCACGCTGTTATCCCTAAAGTAACTTATTT  
 CexTE ATTTGGACTTTCAAATATATTACGCTGTTATCCCTAAAGTAACTTATTT

CioLC1 AAGCTTCAAAAATTTGGGTATCAAATAATACTATTT-TAATACTTTGA  
 CioEC1 AAGCTTCAAAAATTTGGGTATCAAATAATACTATTT-TAATACTTTGA  
 CioMO1 AAGCTTCAAAAATTTGGGTATCAAATAATACTATTT-TAATACTTTGA  
 CioTI1 AAGCTTCAAAAATTTGGGTATCAAATAGTACTATTT-TGATACTTTGA

CiizU1 AAGCTTCAAAAAGTTTTGGGTATCAAATAATGTTATCT-TAATATTATGA  
 CltFB1 AAAGTTCAAAAATTTTTGGGTATTTAAAATAATATTATTT-TAATGTTTTAG  
 CltFB2 AAAGTTCAAAAATTTTTGGGTATTTAAAATAATATTATTT-TAATGTTTTAG  
 Cl1CH1 AAGCTTCAAAAATTTTTGGGTATTTAAAATGATGATATTT-TAATATTGTAA  
 Cl1CH2 AAGCTTCAAAAATTTTTGGGTATTTAAAATGATGATATTT-TAATATTGTAA  
 Cl1TO1 AAGCTTCAAAAATTTTTGGGTATTTAAAATGATGGTATCT-TAATATTGTAA  
 Cl1TZ1 AATCTTCAAAAATTTTTGGGTATTTAAAATAATGATATTT-TAATATTATAG  
 Cl1AR1 AATCTTCAAAAATTTTTGGGTATTTAAAATAATGATATTT-TAATATTATAG  
 Cl1QU1 AAGCTTCAAAAATTTTTGGGTATTTAAAATAATCTATTT-TAGCCCTCTAA  
 Cl1HU1 AAATTTCAAAAATTTTTGGGTATTTAAAATAGTTTTATTT-TAGCCCTCTAA  
 CbaDO AATATTCAAAAAGTTTTGGGTATTTAAACTATCCTATTATAATAATTTAA  
 CviAR AAATTTCAAAAATTTTTGGGTATTTAAAATAATATTATTTTTAATA-TTTAA  
 CexTE AATTTTCAAAAATTTTTGGGTATTTAAAATAATGTCATTTTTA-TATTTTGA

CioLC1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACACACTTTTTAATTTACTA  
 CioEC1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACACACTTTTTAATTTACTA  
 CioMO1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACACACTTTTTAATTTACTA  
 CioTI1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACACACTTTTTAATTTACTA  
 CiizU1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACATACTTTTTAATTTATTA  
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 Cl1CH1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACATACTTTTTAATTTATTA  
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 Cl1TZ1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACATACTTTTTAATTTATTA  
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 Cl1HU1 AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACATACTTTTTAATTTATTA  
 CbaDO AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACATACTTTTTAATTTATTA  
 CviAR AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACATACTTTTTAATTTATTA  
 CexTE AAGTGTTC-ATCTTCCACCGCCCCAGTAAAACATACTTTTTAATTTATTA

CioLC1 AATTATTATATGTAAAGCTTTATAGGGTCTTCTTGTCTAAAAGAGGTATT  
 CioEC1 AATTATTATATGTAAAGCTTTATAGGGTCTTCTTGTCTAAAAGAGGTATT  
 CioMO1 AATTATTATATGTAAAGCTTTATAGGGTCTTCTTGTCTAAAAGAGGTATT  
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 CbaDO GATTATTATATGTAAAGCTTTATAGGGTCTTCTTGTCTAAAAGAACATT  
 CviAR AATTTTCTTATGTAAAGTTTTATAGGGTCTTCTTGTCTAAAAGAACATT  
 CexTE AATTATTTTATGTAAAGCTTTATAGGGTCTTCTTGTCTAAAAGAACATT

CioLC1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 CioEC1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 CioMO1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 CioTI1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 CiizU1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 CltFB1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 CltFB2 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 Cl1CH1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 Cl1CH2 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 Cl1TO1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 Cl1TZ1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA  
 Cl1AR1 TTAGCCTTTTTACTAAAAGGTAATTTTTGAAGAAAAAGCTAAGAAAGAA

Cl1QU1 TTAGCCTTTTTACTAAAAAGTAAAATTCAAAGGAAAACTAAGAAAGAA  
 Cl1HU1 TTAGCCTTTTTACTAAAAAGTAAAATTCAAAGGAAAACTAAGAAAGAA  
 CbaDO TTAGCCTTTTTACTAAAAAGTAAAGTTAAAAAGTAA-AATTA AAAAGAG  
 CviAR TTAGCCTTCTTACTAAAAAGGTGAAATTTGAAATTAATAGTTAAGAAAGAA  
 CexTE TTAGCCTTTTTACTAAAAAGTAAAGTTCAAAGAAAGACCAAGAAAGAA

CioLC1 ACTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 CioEC1 ACTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 CioMO1 ACTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 CioTI1 ACTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 CiiZU1 ACTTCTAGTTTATCCTTTCATTCCAGTCTTAAATTACAAGACTAATGAT  
 CltFB1 ACTCTCCAGTTTATCCTTTCATTCCAGTCTTAAATTACAAGACTAATGAT  
 CltFB2 ACTCTCCAGTTTATCCTTTCATTCCAGTCTTGAATTACAAGACTAATGAT  
 Cl1CH1 GCTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 Cl1CH2 GCTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 Cl1TO1 GCTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 Cl1TZ1 GCTCTCTAGTTTACCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 Cl1AR1 GCTCTCTAGTTTACCCTTTCATTCCAGTCTTAAATTATAAGACTAATGAT  
 Cl1QU1 ACTTCTGGTTTATCCCTTCATCCAGTCTTAAATTATAAGACTAATGAT  
 Cl1HU1 ACTTCTGGTTTACCCTTCATCCAGTCTTAAATTATAAGACTAATGAT  
 CbaDO CTTTTCCAGTCTATCCTTTCATTCCAGTCTTCAATTATAAGACTAATGAT  
 CviAR ATTTTCTAGTTTACCCTTTCATTCCAGTCTTAAATTACAAGACTAATGAT  
 CexTE ACTCTCTAGTTTATCCTTTCATTCCAGTCTTAAATTACAAGACTAATGAT

CioLC1 TATGCTACCTTTGCNACANNNNNN  
 CioEC1 TATGCTACCTTTGCNACACNNNNN  
 CioMO1 TATGCTCTCTTTTNGGCCACANNN  
 CioTI1 TATGCTACCTTTGCNACANNNNNN  
 CiiZU1 TATGCTACCTTTGCNACANNNNNN  
 CltFB1 TATGCTACCTTTGCNNNNNNNNNN  
 CltFB2 TATGCTCTTTTTNGNNCACANNNN  
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 Cl1CH2 TATGCTCTTTTTTGNGCACANNNN  
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 Cl1QU1 TATGCTACCTTTGCNACANNNNNN  
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 CbaDO TATGCTACCTTTGCNACACNNCCN  
 CviAR TATGCTACCTTTGCNACANNNCTN  
 CexTE TATGCTACCTTTGCNACANNNNNN